50 Slide kit contains (Cat. No. 93-3943)		
Reagent 1A	Trypsin Concentrate	3 mL
Reagent 1B	Trypsin Diluent	12 mL
Reagent 2	Denaturing Solution (ready-to-use)	6 mL
Reagent 3	Blocking Solution (ready-to-use)	6 mL
Reagent 4	Biotinylated Ms x BrdU (ready-to-use)	6 mL
Reagent 5	Streptavidin-peroxidase (ready-to-use)	6 mL
Reagent 6A	Substrate Buffer Concentrate (20x)	2 mL
Reagent 6B	DAB Concentrate (20x)	2 mL
Reagent 6C	0.6% Hydrogen Peroxide Concentrate (20x)	2 mL
Reagent 7	Hematoxylin (Ready-to-use)	6 mL
Reagent 8	Histomount™ (ready-to-use)	6 mL
Positive Control	Four unstained BrdU control slides. (Methacarn fixed, paraffin-embedded)	
250 Slide kit contains (Cat. No. 93-3944)		
Reagent 1A	Trypsin Concentrate	15 mL
Reagent 1B	Trypsin Diluent	60 mL
Reagent 2	Denaturing Solution (ready-to-use)	30 mL
Reagent 3	Blocking Solution (ready-to-use)	30 mL
Reagent 4	Biotinylated Ms x BrdU (ready-to-use)	30 mL
Reagent 5	Streptavidin-peroxidase (ready-to-use)	30 mL
Reagent 6A	Substrate Buffer Concentrate (20x)	10 mL

DAB Concentrate (20x)

Hematoxylin (ready-to-use)

Histomount[™] (ready-to-use)

Reagents and materials required but not provided:

Quenching solution for endogenous peroxidase

Phosphate Buffered Saline (PBS)

Handling, storing, and shelf life

Ten unstained BrdU control slides

(Methacarn fixed, paraffin-embedded)

Store kit at 2–8°C. All performance guarantees are void after kit expiration date. Observe necessary health and safety pre-

cautions when using this product. Avoid contact with skin and clothes. Wearing of latex or rubber gloves is recommended.

There is a potential hazard of explosion due to the reaction of

sodium azide, a preservative, with copper metal in plumbing

systems. To avoid this, flush the drain thoroughly with water af-

0.6% Hydrogen Peroxide Concentrate (20x)

Reagent 6B

Reagent 6C

Reagent 7

Reagent 8

Positive Control

Alcohol and xylene

ter disposal of reagents.

Cover slips

Conditions for use

Invitrogen BrdU staining kit is designed for research use only and is not intended for therapeutic or diagnostic purposes.

Expected staining results

BrdU labeling by IHC is specifically exhibited in the nuclei of cells in BrdUincorporated tissue sections. The use of the included positive control slides is recommended. The images below of BrdU labeling by IHC also serve as a reference.



Mouse small intestine stained by immunohistochemistry using the Invitrogen BrdU Staining Kit. Note the specific nuclear labeling by DAB. (A) 5X magnification (B) 20X magnification.

Related products

Invitrogen also supplies a BrdU labeling reagent with recommended procedures (Cat. No. 00-0103) and BrdU control slides (Cat. No. 09-0042).

Reference

10 mL

10 mL

30 mL

25 mL

Ellwart E, Dormer, P (1985) Cytometry 6:513-520. Effect of 5-Fluoro-2'-Deoxyuridine (FdUrd) on 5-Bromo- 2'-Deoxyuridine (BrdUrd) incorporation into DNA measured with monoclonal BrdUrd antibody and by the BrdUrd/Hoechst Quenching Effect.

HistoGrip[™] and Histomount[™] are trademarks of Invitrogen.

Invitrogen BrdU Staining Kit

Streptavidin–Biotin System for BrdU Staining

Cat. No.	Sufficient for
93-3943	50 Slides
93-3944	250 Slides

Introduction

In the past, cell-proliferation studies used radioactive thymidine as an incorporated label to give evidence of DNA replication. The Invitrogen BrdU staining kit uses a less hazardous technique by using bromodeoxyuridine (BrdU), a thymidine analog, in lieu of a radioactive reagent.(1) BrdU is incorporated into proliferating cells (Sphase) much in the same way as radioactive thymidine, but is then detected by a monoclonal anti-BrdU antibody, and revealed using a highly sensitive streptavidin-biotin staining system. Invitrogen BrdU Staining Kit uses a biotinylated monoclonal anti-BrdU, thus eliminating the need for a species-specific secondary antibody. As a result, BrdU labeling can be performed in rats and mice without problems of cross-reactivity or spurious staining.

For research use only. CAUTION: Not intended for human or animal therapeutic or diagnostic use.



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invitrogen

PIN: 30223 Rev. 10/09 DCC-09-1440

Suggested staining procedures

A. For paraffin-embedded tissues

Preparation of slides:

- 1. Tissues should be labeled with BrdU (Invitrogen provides BrdU labeling reagent with complete instructions, Cat. No. 00-0103).
- 2. Fix target tissue in 10% NBF (Neutral Buffered Formalin), or in an alcohol-based fixative (such as alcohol or Methacarn). Process tissues for paraffin embedding.
- Cut tissue 3-4 microns thick and place on HistoGrip™ (Cat. No. 00-8050) or poly-I-lysine coated slides. Dry slides in a 60°C oven for 30-60 minutes.
- Deparaffinize slides in 2 changes of xylene for 5 minutes each. Rehydrate slides in a series of graded alcohol. Slides are now ready for BrdU staining.
- 5. Formalin-fixed tissues require trypsin digestion.

Optional:

6. If alcohol-fixed tissues are used, tissue sections can be postdipped in 10% NBF for 30-60 seconds. Rinse well. This may improve morphology.

For cultured cells and cell suspensions:

Preparation of cells:

- 1. Remove labeling medium from cells and wash in several changes of PBS.
- 2. Fix cells in 70% alcohol or acid-ethanol for 15–30 minutes at 4°C. (Acetone or Methacarn fixatives also can be used.
- **3.** If necessary, block for endogenous peroxidase activity with 3% hydrogen peroxide in methanol for 10 minutes.
- 4. Wash in 3 changes of distilled water for 2 minutes each.

Troubleshooting

• Possible causes of negative or poor staining:

- 1. BrdU labeling was not adequate.
- 2. Staining steps were not performed correctly, or were omitted.
- 3. Longer incubation of biotinylated antibody may be required.
- 4. Tissue, if fixed in formalin, may need further digestion.

• Possible causes for high background staining:

- 1. Tissue may require a longer blocking step.
- 2. Inadequate rinsing of slides.
- 3. Deparaffinization was not complete.
- 4. Over-development of substrate may have occurred.

Reagent Preparation	Staining procedures	Incubation (min)
Peroxidase Quenching Solution: Add one part 30% $\rm H_2O_2$ to 9 parts absolute methanol. Mix well.	Submerge slides in Quenching solution. After incubation, rinse with PBS (2 min, 3 times)	10
Trypsin*: Do Not Digest Control Slides: Add 1 drop of reagent 1A to 3 drops off reagent 1B. Mix well.	For formalin-fixed tissue only (Not necessary for alcohol fixed tissues). Add 2 or more drops to each section. Incubate in moist chamber at 37°C. Rinse in distilled water (2 min, 3 times)	3–10
Do Not Digest Control Slides.		
Denaturing Solution**: Reagent 2 (ready-to-use)	Apply 2 drops or more to each section. Incubate at room temperature. Rinse with PBS (2 min., 3 times)	20–30
Blocking Solution: Reagent 3 (ready-to-use)	Apply 2 drops or 100 μL to each section. Incubate at room temperature. Drain or blot off the solution. Do not rinse.	10
Biotinylated Mouse Anti-BrdU: Reagent 4 (ready-to-use).	Apply 2 drops or 100 μL to each section. Incubate at room temperature. Rinse with PBS (2 min, 3 times)	30–60
Streptavidin-Peroxidase: Reagent 5 (ready-to-use).	Apply 2 drops or 100 μL to each section. Incubate at room temperature. Rinse with PBS (2 min, 3 times)	10
Ab mixture: Add 1 drop of reagents 6A, 6B, and 6C to 1 mL distilled water. Mix well. Protect from light and usewithin one hour.	Apply 2 or more drops of Ab mixture to each section. Rinse well with distilled water.	2–5
Hematoxylin: Reagent 7 (ready-to-use) .	Counterstain the slides with 2 drops or 100 μ L of Hemotoxylin. Wash slides in tap water. Put slides into PBS until sections turn blue (approx. 30 sec). Rinse in distilled water.	1–5
Histomount™: Reagent 8 (ready-to-use).	Dehydrate slides in a graded series of alcohol, and clear with xylene. Add 2 drops of Histomount™ and coverslip.	

* Depending upon the fixative condition, concentration of Trypsin may be adjusted. Please note that we have not characterized this kit using fresh frozen tissue. For fresh frozen tissue, the trypsin step may need to be omitted or performed for 0–1 min.

** Corrosive reagent, handle with caution.

Reagent Preparation	Staining Procedures	Incubation (min)
Dilute Denaturing Solution**: (Reagent 2) 1:1 with distilled water.	Incubate with Denaturing solution. After incubation, rinse in PBS (2 min, 3 times)	30
Blocking Solution: Reagent 3 (ready-to-use).	Apply sufficient quantity to cover specimen. Incubate at room temperature. Drain or blot off the solution. Do not rinse.	10
Biotinylated Mouse Anti-BrdU: Reagent 4 (ready-to-use).	Add sufficient antibody to cover specimen. Incubate at room temperature. Rinse with PBS (2 min, 3 times).	30–60
Streptavidin-Peroxidase: Reagent 5 (ready-to-use).	Add sufficient antibody to cover specimen. Incubate at room temperature. Rinse with PBS (2 min, 3 times).	10
Ab mixture: Add 1 drop of reagents 6A, 6B, and 6C to 1 mL distilled water. Mix well. Protect from light and use within one hour.	Add sufficient quantity of the Ab mixture to cover specimen. Incubate. Rinse well with distilled water.	2–5
Hematoxylin: Reagent 7 (Ready-to-use).	Counterstain with sufficient quantity of Hematoxylin to cover specimen. Wash slides in tap water. Put slides into PBS until sections turn blue (approx. 30 sec). Rinse in distilled water.	1–2
Histomount™: Reagent 8 (Ready-to-use).	Dehydrate slides in a graded series of alcohol, and clear with xylene. Add 2 drops of Histomount [™] and coverslip.	

** Corrosive reagent, handle with caution.

